Determination Of Polycyclic Aromatic Hydrocarbons (PAHs) In Smoked Clarias Gariepinus And Mugil Cephalus From Bayelsa State, Nigeria.

Olubunmi Olusoga Ezomoh And Gwegwe Joyful Sueze

Department Of Biochemistry, Faculty Of Basic Medical Sciences, Niger Delta University.

Abstract

This study evaluated PAH concentrations in smoked Clarias gariepinus and Mugil cephalus obtained from Nembe River, Bayelsa State. Samples were dehydrated in an oven, pulverized, and precisely weighed before n-hexane extraction. Following filtration, the extracts were analyzed using GC-FID to quantify PAH content. The following results were obtained, (6.0 mg/L, 19.81 mg/L), acenaphthylene (7.0 mg/L, 7.0mg/L), acenaphthene (7.0 mg/L, 7.0mg/L), fluorene (9.00 mg/L, 9.0 mg/L), phenanthrene (7.0 mg/L, 7.0 mg/L), anthracene (7.0 mg/L, 7.0mg/L), fluranthene (7.0 mg/L, 7.0mg/L), pyrene (7.0 mg/L, 7.0mg/L), benzo(c)pyrene (7.0 mg/L, 7.0mg/L), benzo(a)anthrancene, (7.0 mg/L, 7.0mg/L), Chrysene (7.0 mg/L, 7.0mg/L), benzo(k)fluoranthene (7.0 mg/L, 7.0mg/L), benzo(e)pyrene (8.0 mg/L, 1.0mg/L), benzo(a)pyrene (7.0 mg/L, 8.0mg/L), 3-methcarboxy anthracene (3.0 mg/L, 3.0 mg/L), benzo(g, h, i)perylene (6.80 mg/L, 7.30 mg/L), dibenz(a, l)perylene (60.3 mg/L, 141.63 mg/L). While DahA was not detected. Results showed that the detected PAHs originated from the fish smoking process. The total PAH concentrations were substantially lower than the maximum limits established by European Union (EU) regulations, Notably, benzo[a]pyrene levels (below 5 ug/kg) complied with EU safety standards, indicating minimal carcinogenic risk and confirming the smoked fish products' safety for human consumption.

Keywords: Smoked fish, Clarias gariepinus, Mugil cephalus, GC-FID, GC/MS, Polycyclic aromatic hydrocarbons (PAHs), Bayelsa state.

Date of Submission: 05-08-2025 Date of Acceptance: 15-08-2025

I. Introduction

PAHs represent a class of organic molecules containing multiple condensed aromatic ring structures. These persistent compounds are ubiquitously distributed in environmental matrices (water, air, soil) and occur in trace amounts in various food products. Thermal food processing techniques including smoking, grilling, frying and baking serve as primary contamination pathways. Most PAHs originate as environmental pollutants from incomplete organic matter combustion during industrial and anthropogenic activities (Ishizaki et al., 2021). Recognized for their carcinogenic potential, PAHs are designated as priority contaminants under EU and USEPA regulations. Epidemiological data indicate human exposure accounts for 58-98% of PAH contamination incidents (Farhadian et al., 2021).

Polycyclic aromatic hydrocarbons are systematically classified into two primary categories according to their molecular properties: high molecular weight (HMW) and low molecular weight (LMW) PAHs, with this division reflecting significant differences in their chemical behavior and biological interactions. HMW PAHs contain 4-6 aromatic rings, exhibit slower biodegradation by native microbes, and tend to persist in aquatic systems. Due to their bioaccumulation potential in marine life (e.g., fish and mussels) and heightened carcinogenicity, they pose significant environmental risks (Rocher et al., 2024). PAHs with low molecular weight (LMW), comprising 2-3 aromatic ring structures, demonstrate significantly reduced carcinogenic potential when compared to high molecular weight (HMW) PAHs. However, they still demonstrate significant toxicity to numerous aquatic species, making them environmentally concerning. (Brown et al., 2018).

Smoking represents one of the most traditional techniques for fish preservation and processing. This method involves exposing fish to smoke generated from burning wood or plant matter. The process typically incorporates simultaneous salting, drying, heating and smoking within a specialized chamber. Its preservative effects primarily stem from moderate dehydration and the deposition of both aliphatic and aromatic compounds on the fish's surface (Simpko, 2017). High-temperature food preparation techniques including smoking, drying, roasting, baking and frying are well-established as significant contributors to PAH contamination in food products (Yurchenko and Mölder, 2019).

Wood smoke comprises numerous PAHs and their derivatives, including carcinogenic compounds like Benzo[a]Pyrene (BaP). BaP serves as the primary indicator for carcinogenic PAHs in smoked fish, with a regulatory limit of $2\mu g/kg$. Following metabolic conversion to diol-epoxides in mammalian cells, PAHs form covalent bonds with DNA and other macromolecules. This interaction induces DNA replication errors and mutations, initiating carcinogenesis a activation pathway shared, with minor variations, by all carcinogenic PAHs. (Falco *et al.*, 2016).

This study primarily aimed to quantify the levels and assess the potential health risks associated with polycyclic aromatic hydrocarbons (PAHs) in smoked mullet and river catfish commonly consumed in Bayelsa State, Nigeria.

II. Materials

The study utilized smoked African catfish (*Clarias gariepinus*) and flathead mullet (*Mugil cephalus*) samples. Laboratory equipment included a drying oven, spatula, homogenization tools (mortar and pestle) and analytical balance. Extraction procedures utilized n-hexane and dichloromethane with extraction bottles and filter paper. Quantitative analysis was conducted using an American-manufactured HP 5890 Series II gas chromatograph with flame ionization detection (GC-FID).

III. Methods

Extraction of Fish Sample

Smoked *Clarias gariepinus and Mugil cephalus* was purchased from a road side vendor at Basambiri, Nembe, while smoked ones was purchased from Swali market in Yenagoa, Bayelsa State. The purchased smoked fishes were chopped finely and homogenized, after which bone fragments were removed using tweezers. The fish sample were subjected to oven drying at 60°C for 48 hours. Following dehydration, the samples were removed and pulverized. A precisely weighed 1g aliquot of homogenized fish material was measured into an extraction vessel, to which 10 ml of n-hexane was added. After a 72-hour standing period, the solution was vacuum-filtered through Whatman filter paper, the clarified extract was then subjected to GC-FID analysis for PAH quantification.

Gas Chromatography-Flame Ionization Detector (GC-FID) Analysis

Chromatographic separation was conducted using an HP 5890N Series II GC system configured with split/splitless injection capability and FID detection. An Agilent HP-608 capillary column (30 m length \times 0.53 mm internal diameter, 0.5 µm stationary phase thickness) achieved compound resolution. The thermal profile began at 70°C (2 min isothermal), increased at 6°C/min to 260°C (15 min final hold), with injector and detector maintained at 280°C and 300°C respectively. Nitrogen served as carrier gas at 4.0 mL/min constant flow rate. ChemStation software controlled instrument operation and data processing. PAH identification involved retention time matching with pure standards, with GC/MS confirmation for samples showing elevated PAH levels.

IV. Results And Data Presentation

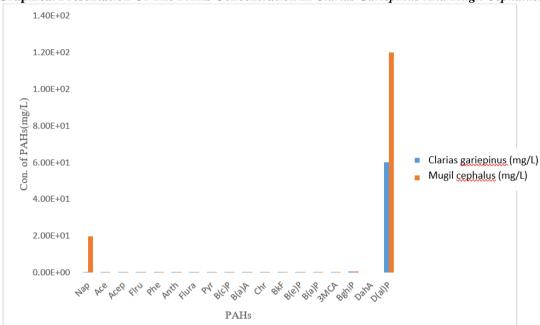
Table 1: Level of PAHs in Clarias gariepinus and Mugil cephalus

Permissible limits of PAHs in Clarias gariepinus and Mugil cephalus by international bodies.

PAHs	Clarias gariepinus	Mugil	Permissive	
	(mg/Kg)	cephalus	Limit(mg/kg)	Source
		(mg/Kg)		
Nap	6. 0E-02	19.81		
Ace	7. 0E-02	7. 0E-02		
Acep	7. 0E-02	7.0E-02	0.06	USEPA
Flru	9. 0E-02	9. 0E-02	0.06	SON
Phe	7. 0E-02	7. 0E-02	0.1	USEPA
Anth	7. 0E-02	7. 0E-02	0.3	USEPA
Flura	7. 0E-02	7.0E-02		
Pyr	7. 0E-02	7. 0E-02	0.06	SON
B(c)P	7. 0E-02	7. 0E-02	3mg/kg	WHO
B(a)A	7. 0E-02	7. 0E-02	10mg/kg	WHO
Chr	7.0E-02	7. 0E-02	10mg/kg	WHO
BkF	7. 0E-02	7. 0E-02	0.10	USEPA
B(e)P	8. 0E-02	1. 0E-01	5.10	USEPA
B(a)P	7. 0E-02	8. 0E-02		
3MCA	3. 0E-02	3. 0E-02		
BghiP	6. 8E-01	7. 30E-01	0.05	SON
DahA	0.001	0	0.03	SON
D(a,l)P	60.03	120.09		

Table 1 shows the quantitative analysis results for PAH present in both *Clarias gariepinus* and *Mugil cephalus* with recommended values by standard organizations. The predominant PAHs found in the smoked fishes are Nap, D(a,i)P. Although, the two PAHs are not among biomarkers of PAHs that are dangerous to human health

but continual consumption might result in bioaccumulation in the body of the consumers which may cause ill health.



Graphical Presentation Of The PAHs Concentration In Clarias Gariepinus And Mugil Cephalus.

Table 1: Level of PAHs in CLARIAS GARIEPINUS AND MUGIL CEPHALUS

PAHs	Clarias gariepinus (mg/L)	Mugil cephalus(mg/L)
Nap	6.00E-02	19.81
Ace	7.00E-02	7.00E-02
Acep	7.00E-02	7.00E-02
Flru	9.00E-02	9.00E-02
Phe	7.00E-02	7.00E-02
Anth	7.00E-02	7.00E-02
Flura	7.00E-02	7.00E-02
Pyr	7.00E-02	7.00E-02
B(c)P	7.00E-02	7.00E-02
B(a)A	7.00E-02	7.00E-02
Chr	7.00E-02	7.00E-02
BkF	7.00E-02	7.00E-02
B(e)P	8.00E-02	1.00E-01
B(a)P	7.00E-02	8.00E-02
3MCA	3.00E-02	3.00E-02
BghiP	6.80E-01	7.30E-01
DahA	0	0
D(a,l)P	60.03	120.09
Sum	6.17E+01	141.63
Mean	3.43	7.868333
Standard deviation	±0.33	±0.669

From **Table 1**: It shows the level of PAHs in my sample with recommended values by standard organization. The predominant PAHs found in the smoked fishes are Nap, D (a,i)P. Although, the two PAHs are not among biomarkers of PAHs that are dangerous to human health but continual consumption might result in bioaccumulation in the body of the consumers which may cause ill health.

Table 2 Pearson's Correlation interpretation.

S/No	Degree of Correlation	Types of Correlation	
1	± 0.00 to ± 0.20	Negligible	
2	±0.20 to ±0.40	Low	
3	$\pm 0.40 \text{ to} \pm 0.70$	Moderate	
4	$\pm 0.70 \text{ to} \pm 0.90$	High	
5	±0.90 ±1.00	Very High	
6	±1.00	Perfect	

Table 3: Interrelationship between the analyzed PAHs in the selected Fishes samples from Bayelsa State

	Clarias gariepinus	Mugil cephalus
Clarias gariepinus	1	
Mugil cephalus	0.986448	1

Interrelationship between the analyzed PAHs in the selected Fishes samples from Bayelsa State

During this study, high correlations between *Clarias gariepinus* and *Mugil cephalus* (r = 0.986448), was observed in samples of selected smoked fishes. Hence, correlations among the Fishes show that the analyzed sample (*Clarias gariepinus and Mugil cephalus*) had sources of PAHs from the same point/origin. The consumers of these smoked fishes will be obtaining PAHs in their diet, which will bioaccumulated in their body and had adverse effect in their well-being and general health conditions.

t-Test: Two-Sample Assuming Equal Variances				
	Clarias gariepinus	Mugil cephalus		
Mean	3.43	7.868333		
Variance	199.5511	805.8976		
Observations	18	18		
Pooled Variance	502.7243	0.986448		
Hypothesized Mean Difference	0	0		
df	34	17		
t Stat	-0.59385	1.286381		
P(T<=t) one-tail	0.278272	0.107777		
t Critical one-tail	1.690924	1.739607		
P(T<=t) two-tail	0.556544	0.215554		
t Critical two-tail	2.032245	2.109816		

V. Discussion

The data in table 1 shows that the analyzed samples contain 17 varieties of polycyclic aromatic hydrocarbons including naphthalene (6.0 mg/L, 19.81 mg/L), acenaphthylene (7.0 mg/L, 7.0mg/L), acenaphthene (7.0 mg/L, 7.0mg/L), fluorene (9.00 mg/L, 9.0 mg/L), phenanthrene (7.0 mg/L, 7.0 mg/L), anthracene (7.0 mg/L, 7.0mg/L), fluranthene (7.0 mg/L, 7.0mg/L), pyrene (7.0 mg/L, 7.0mg/L), benzo(c)pyrene (7.0 mg/L, 7.0mg/L), benzo(a)anthracene, (7.0 mg/L, 7.0mg/L), Chrysene (7.0 mg/L, 7.0mg/L), benzo(k)fluoranthene (7.0 mg/L, 7.0mg/L), benzo(e)pyrene (8.0 mg/L, 1.0mg/L), benzo(a)pyrene (7.0 mg/L, 8.0mg/L), 3-methcarboxy anthracene (3.0 mg/L, 3.0mg/L), benzo(g, h, i)perylene (6.80 mg/L, 7.30mg/L), dibenz(a, l)perylene (60.3 mg/L, 141.63mg/L). These findings corroborate previous studies demonstrating that while raw food products typically contain minimal PAH concentrations, these hazardous compounds primarily form during thermal processing techniques including smoking, roasting, baking and frying (Kayali *et al.*, 2019). Within this group of priority PAHs, five compounds exhibit carcinogenic properties: chrysene, benzo[a]pyrene, benzo[k]fluoranthene, benzo[g,h,i]perylene and benz[a]anthracene. In contrast, fluorene, acenaphthylene, pyrene, acenaphthene, phenanthrene, naphthalene, anthracene and fluoranthene are classified as non-carcinogenic (Kafeelah *et al.*, 2019).

High significant concentration of different polycyclic aromatic hydrocarbons were observed in samples of selected smoked fishes. Hence, correlations among the fishes show that the analyzed samples smoked (*Clarias gariepinus and Mugil cephalus*) had sources of PAHs from the same point/origin. For the *Clarias gariepinus*, the concentrations ranged from 6.0 mg/L for Nap, to 60.03 mg/L for (a,i)P. On the other hand, the concentrations of PAHs in the *Mugil cephalus* ranged from 1.0 mg/L for B(e)P to 120.09 mg/L D(a,l)P. The consumers of these smoked fishes will be obtaining PAHs in their diet, which may accumulate in their body and cause adverse effects that will affect their well-being and general health conditions.

VI. Conclusion

Analysis revealed that PAH concentrations in smoked *Clarias gariepinus* and *Mugil cephalus* samples fell below the safety thresholds established by the European Commission, FAO, and WHO. These low levels suggest the smoked fish products are safe for human consumption and unlikely to present carcinogenic or other health risks to Bayelsa State residents or other consumers. The detected PAHs were predominantly pyrogenic in origin, resulting from the smoking process itself.

Acknowledgement

The authors wish to thank Dr. Vaikosen E. Nicholas, Head of Department, Department of Pharmaceutical and Medicinal Chemistry, Faculty of Pharmacy and Dr. Chukwudi Njoku, Head of Department, Department of Pharmacology, Faculty of Basic Clinical Sciences, Niger Delta University, for allowing us use the department's laboratory for this study. We also wish to extend our gratitude to Dr. Raji Rafiu, Lab Technologist, Department of Pharmaceutical and Medicinal Chemistry, Faculty of Pharmacy, Niger Delta University for providing the available technical knowledge and support for the study.

References

- [1] Alexandrov, K., Rojas, M., & Satarug, S. (2015). The Critical DNA Damage By Benzo (A) Pyrene In Lung Tissues Of Smokers And Approaches To Preventing Its Formation. Toxicology Letters, 198(1), 63-68.
- [2] Brown, J. And Peake, B. (2018). "Sources Of Heavy Metals And Polycyclic Aromatic Hydrocarbons In Urban Storm Water Runoff." Sci. Total Environ, Vol. 359, Pp. 145-155.
- [3] Directive, C. (2023). 107/EC Of The European Parliament And Of The Council Of 15 December 2004 Relating To Arsenic, Cadmium, Mercury, Nickel And Polycyclic Aromatic Hydrocarbons In Ambient Air. From The Official Journal Of The European Communities, 26(2005).
- [4] Downward, G. S., Hu, W., Rothman, N., Reiss, B., Wu, G., Wei, F., Chapman, R. S., Portengen, L., Qing, L., & Vermeulen, R. (2022). Polycyclic Aromatic Hydrocarbon Exposure In Household Air Pollution From Solid Fuel Combustion Among The Female Population Of Xuanwei And Fuyuan Counties, China. Environmental Science & Technology, 48(24), 14632–14641.
- [5] Falcó, G., Domingo, J. L., Llobet, J. M., Teixidó, A., Casas, C., & Müller, L. (2016). Polycyclic Aromatic Hydrocarbons In Foods: Human Exposure Through The Diet In Catalonia, Spain. Journal Of Food Protection, 66(12), 2325–2331. https://Doi.Org/10.4315/0362-028x-66.12.2325
- [6] Farhadian, A., Jinap, S., Han, F. H. N., And Zaidual, I. S., (2021). "Effects Of Meat Preheating And Wrapping On The Levels Of Polycyclic Aromatic Hydrocarbons In Charcoal Grille Meant. Food Chemical, 12, 141-146.
- [7] Idowu, O., Semple, K. T., Ramadass, K., O'Connor, W., Hansbro, P., & Thavamani, P. (2019). Beyond The Obvious: Environmental Health Implications Of Polar Polycyclic Aromatic Hydrocarbons. Environment International, 123, 543–557. https://Doi.Org/10.1016/J.Envint.2018.12.051
- [8] Ishizaki, A., Saitoa, K., Haniokab, N., Narimatsub, S., And Kataokaa, H. (2021). Determination Of Polycyclic Aromatic Hydrocarbons In Food Samples By Automated On-Line In-Tute Solid-Phase Microexraction Coupled With High-Performance Liquid Chromatography-Fluorescence Detection. Journal Of Chromatogram, 1217, 5555-5563
- [9] Käfferlein, H. U., Marczynski, B., Mensing, T., & Brüning, T. (2010). Albumin And Hemoglobin Adducts Of Benzo [A] Pyrene In Humans—Analytical Methods, Exposure Assessment, And Recommendations For Future Directions. Critical Reviews In Toxicology, 40(2), 126-150.
- [10] Kayali, S. M. N., Rubio-Barroso, S., Cuesta-Jimwnez, M. P., And Polo-Diez, L. M. (2019). A New Method For The Determination Of Selected Pahs In Coffee Brew Samples By HPLC With Fluorimetric Detection And Solid-Phase Extraction. Journal Of Liquid Chromatography And Related Technology, 22, 615-627.
- [11] Kefeelah, Y. A., Lucy, E. N., Kafayat, F. A., Shehu, A. L., Julius, A. I., And Titus, O. O. (2015). Influence Of Fish Smoking Method On Polycyclic Aromatic Hydrocarbons Contents And It Possible Risks To Human Health. African Journal Of Food Science, 9, 126-135.
- [12] Kondraganti, S. R., Fernandez-Salguero, P., Gonzalez, F. J., Ramos, K. S., Jiang, W., & Moorthy, B. (2023). Polycyclic Aromatic Hydrocarbon-Inducible DNA Adducts: Evidence By 32P-Postlabeling And Use Of Knockout Mice For Ah Receptor-Independent Mechanisms Of Metabolic Activation In Vivo. International Journal Of Cancer, 103(1), 5-11.
- [13] Liu, Y., Vinje, J., Pacifico, C., Natile, G., & Sletten, E. (2022). Formation Of Adenine—N3/Guanine—N7 Cross-Link In The Reaction Of Trans-Oriented Platinum Substrates With Dinucleotides. Journal Of The American Chemical Society, 124(43), 12854-12862.
- [14] Maliszewska-Kordybach, B. (2020). Sources, Concentrations, Fate And Effects Of Polycyclic Aromatic Hydrocarbons (Pahs) In The Environment. Part A: Pahs In Air. Polish Journal Of Environmental Studies, 8, 131-136.
- [15] Mazumdar, S., Redmond, C., Sollecito, W., & Sussman, N. (2016). An Epidemiological Study Of Exposure To Coal Tar Pitch Volatiles Among Coke Oven Workers. Journal Of The Air Pollution Control Association, 25(4), 382-389.
- [16] Orodu, V. E., & Sunny, P. S. (2018). Assessment Of Polycyclic Aromatic Hydrocarbons (Pahs) In Smoked Fish (Hydrocynus Forskahlii) From Tombia Community-Ekpetiama, Bayelsa State. Sumerianz J Biotechnology, 1(4), 100-106.
- [17] Pfeifer, G. P., & Besaratinia, A. (2019). Mutational Spectra Of Human Cancer. Human Genetics, 125, 493-506.
- [18] Rocher, V., Azimi, S., Moilleron, R., And Chebbo, G., (2024). Hydrocarbons And Heavy Metals In The Different Sewer Deposits In The Le Marais Catchment (Paris, France): Stocks, Distributions And Origins. Science Total Environment, 332, 107-122.
- [19] Rybicki, B. A., Nock, N. L., Savera, A. T., Tang, D., & Rundle, A. (2006). Polycyclic Aromatic Hydrocarbon-DNA Adduct Formation In Prostate Carcinogenesis. Cancer Letters, 239(2), 157-167.
- [20] Simko P. (2017). Determination Of Polycyclic Aromatic Hydrocarbons In Smoked Meat Products And Smoke Flavouring Food Additives. Journal Of Chromatography. B, Analytical Technologies In The Biomedical And Life Sciences, 770(1-2), 3–18. https://Doi.Org/10.1016/S0378-4347(01)00438-8
- [21] Stading, R., Gastelum, G., Chu, C., Jiang, W., & Moorthy, B. (2021). Molecular Mechanisms Of Pulmonary Carcinogenesis By Polycyclic Aromatic Hydrocarbons (Pahs): Implications For Human Lung Cancer. Seminars In Cancer Biology, 76, 3-16.
- [22] World Health Organization. (2023). Health Risks Of Persistent Organic Pollutants From Long-Range Transboundary Air Pollution.
- [23] Yabiku, H. Y., Martins, M. S., & Takahashi, M. Y. (2020). Levels Of Benzo [A] Pyrene And Other Polycyclic Aromatic Hydrocarbons In Liquid Smoke Flavour And Some Smoked Foods. Food Additives & Contaminants, 10(4), 399-405.

- [24] Yin, S., Tang, M., Chen, F., Li, T., & Liu, W. (2017). Environmental Exposure To Polycyclic Aromatic Hydrocarbons (Pahs): The Correlation With And Impact On Reproductive Hormones In Umbilical Cord Serum. Environmental Pollution (Barking, Essex :1987), 220(Pt B), 1429–1437.
- [25] Yurchenko, S., & Mölder, U. (2019). The Determination Of Polycyclic Aromatic Hydrocarbons In Smoked Fish By Gas Chromatography Mass Spectrometry With Positive-Ion Chemical Ionization. Journal Of Food Comparative Annals, 18, 857-869.